

1 Cover letter

2 June 16, 2015

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4 To Editor-in-Chief

5 Environmental Health and Preventive Medicine

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7 I, along with my coauthors, would like to ask you to consider the attached manuscript entitled
8 “XPC intron11 C/A polymorphism as a risk factor for prostate cancer” for publication in
9 Environmental Health and Preventive Medicine as an original article.

10

11 This manuscript has not been published or presented elsewhere in part or in entirety, and is
12 not under consideration by another journal. All study participants provided informed consent,
13 and the study design was approved by the appropriate ethics review boards. All the authors
14 have approved the manuscript and agree with submission to your esteemed journal. There are
15 no conflicts of interest to declare.

16

17 Sincerely,

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26 Title page

27 Regular Article

28 Title: XPC intron11 C/A polymorphism as a risk factor for prostate cancer

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44 Brief running title: XPC intron11 C/A polymorphism and prostate cancer

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46 Keyword: Cancer risk, XPC-PAT, DNA repair gene, Xeroderma pigmentosum, Prostate

47 cancer

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51 ABSTRACT

52 Objectives: DNA repair genes play an important role in protection against environmental and
53 endogenous DNA damage, and constitute the first line of defense against cancer. Xeroderma
54 pigmentosum complementation group C (XPC) is involved in the damage recognition step
55 during nucleotide excision repair. The relationship between XPC intron11 C/A polymorphism
56 and cancer risk has not been widely studied. Hence, this study evaluated the relationship
57 between the XPC intron11 C/A polymorphism and prostate cancer risk.

58 Material and Methods: This hospital-based cohort consisted of 152 patients with prostate
59 cancer and 142 male controls. The XPC intron11 C/A genotype was determined using the
60 PCR-RFLP method. Medical, occupational, and cigarette-smoking history was obtained from
61 each participant using questionnaires.

62 Results: Logistic regression analysis revealed that compared to controls, the frequencies of
63 the A/A and C/A genotypes were significantly higher than those of the C/C genotype in
64 cancer patients (OR = 2.03, 95% confidence interval [CI] = 1.03–3.98 and OR = 1.91, 95% CI
65 = 1.13–3.24, respectively). We also found that the frequency of the A/A genotype was
66 significantly higher in cancer cases than in controls among non-smokers (OR = 7.7, 95% CI =
67 1.38–42.88, compared to the C/C genotype).

68 Conclusion: We found that the XPC intron11 C/A polymorphism was associated with an
69 increased risk of prostate cancer. Among non-smokers, the A/A genotype was significantly
70 more prevalent in prostate cancer patients than in controls.

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76 INTRODUCTION

77 Prostate cancer is the most common cancer in men in the United States [1]. However, the
78 incidence of prostate cancer in Asia is relatively low. Risk factors for prostate cancer are diet
79 [2], age, smoking and somatic genomic changes, including deletions, amplifications, and
80 point mutations in tumor suppressor and DNA repair genes [3, 4], similar to those for other
81 cancers.

82 DNA repair genes play an important role in protection against environmental and
83 endogenous DNA damage, and constitute the first line of defense against cancer. The four
84 major pathways of DNA repair are base excision repair, nucleotide excision repair (NER),
85 double strand break repair, and mismatch repair. The xeroderma pigmentosum (XP)
86 complementation group C (XPC) protein is involved in early damage recognition and
87 initiation of NER by binding to HR23B to form the stable XPC-HR23B complex, which
88 recognizes and binds to damaged DNA, leading to subsequent DNA repair [5]. There are
89 three polymorphisms frequently detected in the XPC gene: the poly AT insertion/deletion on
90 intron9 (PAT), the A to C substitution in exon 15 (Lys939Gln), and the C to A substitution in
91 position 5 of intron 11 (intron11 C/A polymorphism).

92 Epidemiologic studies of cancer patients have shown that the PAT+/+ genotype was
93 associated with an increased risk of squamous cell carcinoma of the head and neck [6], and
94 lung cancer risk [7]. The A to C substitution in exon 15 that gave rise to a lysine to glutamine
95 substitution at position 939 has been associated with relatively high risk of bladder cancer [8]
96 and lung cancer [9], but not related to bladder cancer[10]. However, the relationship between
97 the intron11 C/A polymorphism and cancer risk has not been widely studied; to date, the
98 association of the intron11 C/A polymorphism with colorectal cancer, reported by Gil et al,
99 was the only published study [11]. In the present study, we evaluated the risk of prostate
100 cancer associated with the intron11 C/A polymorphism.

101 MATERIALS AND METHODS

102 Subjects

103 The patient consisted of 152 patients with prostate cancer (cases), histologically diagnosed
104 between September 1992 and June 2003 at the University of Occupational and Environmental
105 Health (UOEH) Hospital or the University of Miyazaki Hospital, Japan. The control group
106 consisted of 142 patients with non-cancerous diseases, randomly selected from the UOEH
107 hospital, a hospital near the UOEH Hospital, and the University of Miyazaki Hospital
108 between September 1996 and June 2003. Control patients were examined to rule out
109 urothelial disease, hematuria, and cancer.

110 The demographic data of cases and controls are shown in Table 1. The mean ages were
111 71.7 and 70.2 years for cases and controls, respectively. All study subjects completed a
112 questionnaire administered by a trained interviewer, which covered medical, occupational,
113 and cigarette-smoking history. No exposure to carcinogens, heavy metals, or radiation was
114 recorded in the occupational history of any participant. Cigarette-smoke exposure was
115 calculated as pack-years (1 pack [20 cigarettes]/day X years of smoking). “never-smoker”
116 were defined as those who did not smoke at the time of completing the questionnaire. A
117 “light-smoker” was defined as a person who had smoked less than 35 pack-years, and a
118 “heavy-smoker” was defined as someone who had smoked more than 35 pack-years. “Smoker”
119 in the table 1 included “light-smoker” and “heavy-smoker”. The nature of the study was
120 explained to all participants, and informed consent was obtained from each participant.
121 Ethical approval for the study was obtained from the Ethical Committee of the Faculty of
122 Medicine, University of Miyazaki.

123

124 Genotyping

125 Blood samples were taken from all participants, and genomic DNA was isolated from
126 peripheral leukocytes by proteinase K digestion and phenol/chloroform extraction. The
127 PCR-RFLP method, originally described by Marin et al [7], was used to identify the intron11
128 C/A polymorphism. The PCR primers used for amplification were as follows: forward
129 5'-GCCAAATGCTGACTTGCTCACCGG-3' and reverse
130 5'-GCCACGCGGTGTAGATTGGG-3'. Each 50 μ L PCR reaction mixture contained 10 pmol
131 of each primer, 2.0 mM MgCl₂, 200 mM of each dNTP, 1 unit of Taq polymerase, and 100–
132 300 ng of genomic DNA. The reaction mixture was preincubated for 5 min at 94°C. The PCR
133 conditions used were 30 cycles of 94°C for 30s and 65°C for 30s, followed by 72°C for 30s.
134 The PCR products were digested with the restriction enzyme HaeIII (New England Biolabs,
135 Beverly, MA, USA) at 37°C for overnight. DNA fragments were electrophoresed in 2%
136 agarose gel and stained with ethidium bromide. The A/A genotype gave a single 128 bp band,
137 the C/A genotype showed three bands of 24 bp, 104 bp, and 128 bp, and the C/C genotype
138 had two bands of 24 bp and 104 bp.

139

140 Statistical analysis

141 Univariate analysis was initially performed to compare the distributions of age, sex, and
142 smoking status. Differences in the distributions between cases and controls were tested using
143 the χ^2 and Mann–Whitney U tests, where appropriate. A test for Hardy-Weinberg equilibrium
144 among the controls was conducted using observed genotype frequencies and a χ^2 test. The
145 odds ratio (OR) and 95% confidence intervals (95% CI) for prostate cancer risk were adjusted
146 for age by multiple logistic regression analysis using the Dr SPSS II for Windows (SPSS
147 version11.0.1).

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149 RESULTS

150 The analysis included 152 prostate cancer patients and 142 controls from the Japanese
151 population. The characteristics of the cases and controls, such as age and smoking status, are
152 summarized in Table 1. The prostate cancer patient group had a significantly higher number
153 of never smokers than the control group ($P < 0.01$).

154 The intron11 C/A polymorphism distribution for the cases and controls is shown in Table 2.
155 The distribution of the genotypes among the controls was consistent with the Hardy-Weinberg
156 equilibrium ($P = 0.38$), and was similar to that in another report [7]. The frequencies of the
157 C/A and A/A genotypes were significantly higher in cancer patients than in the control group.
158 The adjusted ORs for prostate cancer risk associated with the C/A and A/A genotypes
159 compared to the C/C genotype were 1.91 (95% CI, 1.13–3.24) and 2.03 (95% CI, 1.03–3.98),
160 respectively.

161 Table 3 shows the distribution of the three genotypes according to smoking status.
162 Thirty-five pack-years was the mean median smoking exposure among the smoking
163 participants in this study. Among never-smokers, the A/A genotype was significantly more
164 frequent in cancer patients than in the controls; the OR was 7.70(95% CI, 1.38–42.88)
165 compared to the C/C genotype.

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167 DISCUSSION

168 This is the first reported study of the intron11 C/A polymorphism in a Japanese population.
169 In this study, we evaluated the association of the intron11 C/A polymorphism with risk of
170 prostate cancer. The genotypic distribution of cases and controls is shown in Table 2. The
171 adjusted ORs for prostate cancer associated with the C/A genotype and A/A genotype
172 compared to C/C genotype were 1.912 (95% CI 1.13–3.24) and 2.03 (95% CI 1.03–3.98),
173 respectively. Similar to the findings of Gil et al in colorectal cancer [11], our results show that
174 the C/A and A/A genotypes are associated with an increased risk of prostate cancer compared
175 to the C/C genotype.

176 XP is a rare recessive disorder associated with a high rate of sunlight-induced skin cancer
177 [12]. XPC is one of seven xeroderma pigmentosum (XP) complementation groups with three
178 common polymorphisms. Several published reports have previously described associations of
179 cancer risk with these XPC polymorphisms [6, 7, 13-18]. Epidemiologic studies of cancer
180 patients have shown an association between the PAT+/+ genotype and a 1.85-fold increase in
181 the occurrence of squamous cell carcinoma of the head and neck [6] and a 1.6-fold increase in
182 the occurrence of lung cancer [7]. However, the relationship between the three XPC
183 polymorphisms and several types of cancer remains controversial [6, 7, 10, 19, 20].

184 Only one study evaluating the relationship between the intron11 C/A polymorphism and
185 colorectal cancer has been reported[11], therefore, the evaluation of the intron11 C/A
186 polymorphism as a risk factor for cancer has not been established. This is only the second
187 report of a relationship between the intron11 C/A polymorphism and cancer risk. The
188 XPC-PAT polymorphism had linkage disequilibrium with XPC exon 15 Lys939Gln
189 polymorphism and intron 11 C/A polymorphism [21]. But the exon 15 Lys939Gln
190 polymorphism didn't change XPC function in vitro [21]. The intron11 C/A polymorphism is a
191 splice acceptor site polymorphism, and is related to an increased frequency of exon 12

192 skipping[7]. The abnormally spliced XPC mRNA iso-form has diminished DNA repair
193 activity and may thereby contribute to cancer susceptibility [22]. The homogenous A/A
194 variant is associated with about 50% reduction of DNA capacity [22]. Our result was
195 supported with these finding.

196 Some studies indicated that the polymorphism of XPC was risk for cancer, but some
197 studies didn't indicate same result. The results of studies concerned XPC polymorphisms was
198 inconsistency. Linkage disequilibrium was reported to the reason for this discrepancy [23].
199 That article indicated that the discrepancy was that the XPC polymorphism evaluated exist in
200 variable degrees of linkage disequilibrium with other that were not evaluated in their
201 investigations [23]. There was no report concerned to linkage disequilibrium in Japanese.
202 Therefore, further study concerned linkage disequilibrium of XPC was needed to evaluation
203 the relation between XPC polymorphism and cancer risk.

204 In general, the distribution of a polymorphism could be changed depending on race.
205 However, there were no reports about the intron 11 C/A polymorphism among Japanese; this
206 report was first article evaluated the intron 11 C/A polymorphism. Our distribution of intron
207 11 C/A polymorphism was the same to other reports [7, 11]. The distribution of the intron 11
208 C/A polymorphism of Japanese couldn't be so different to other results.

209 We also evaluated the association between smoking status and intron11 C/A polymorphism
210 genotype with regard to the risk of prostate cancer. We have shown that the prevalence of the
211 A/A genotype in non-smokers is significantly higher in cancer patients than in controls
212 (adjusted OR = 7.70, 95% CI = 1.38–42.88). Amos et al [24], Khoury et al [25], and Wang et
213 al [26] have also reported that genetic variation associated with cancer risk might be smaller
214 when carcinogen exposure is greater. Jin et al [27] also indicated that the high risk associated
215 with the Pro/Pro genotype of p53 codon 72 polymorphism was associated with lighter
216 smoking. Wang et al. also reported that the same p53 polymorphism was slightly

217 over-represented in lung cancer patients who were non-smokers [26]. There was other report
218 with same result[28]. An explanation for this might be that smoking also alters the level by
219 triggering and up-regulating DNA repair enzymes [29]. Shen et al showed that either
220 inadequate response to DNA damage or inaccurate repair of DNA may have contributed to
221 the risk of lung cancer development in non- or light smokers [30].

222 This was only the second study describing the relationship between the XPC intron11 C/A
223 polymorphism and cancer risk. And our result was first article concerned to prostate cancer.
224 However, in this article there are some limitations. First of all, we collected sample randomly
225 to delete bias. But our sample was small, therefore there could be bias in the sample. Second
226 limitation was more never-smokers in cases than controls. Though smoking could be
227 confounding factor, the reason that cases contained more nonsmokers was that there were
228 many unknown person concerned to smoking status in cases and controls. We evaluated odds
229 ratio by using multiple logistic regression analysis. We thought that the effect of the
230 difference of nonsmokers to the result would be small. The third limitation was that control
231 was hospital control. Hospital control might had some diseases, and the effect of their disease
232 to occurrence of prostate cancer couldn't remove completely. But we excluded persons with
233 urothelial disease, hematuria, and any cancer. More over linkage disequilibrium was needed
234 to evaluate. For these limitations, further evaluation would be needed to confirm the
235 significance of the intron11 C/A polymorphism as a risk factor for prostate cancer.

236 This is the first study reporting that the allele of the intron11 C/A polymorphism of the
237 XPC gene may be a risk factor for prostate cancer in the Japanese population. The prevalence
238 of the A/A genotype in non-smokers was significantly higher in cancer patients than in the
239 controls, and therefore, the A/A genotype may represent a specific cancer risk factor for
240 non-smokers.

241

242 Conflict of interest

243 We acknowledge that we have no conflict of interest.

244

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250 Reference

- 251 1. Jemal A, Kulldorff M, Devesa SS, Hayes RB, Fraumeni JF, Jr. A geographic analysis
252 of prostate cancer mortality in the United States, 1970-89. *Int J Cancer*. 2002;
253 101(2):168-74.
- 254 2. Pienta KJ, Esper PS. Risk factors for prostate cancer. *Annals of internal medicine*.
255 1993; 118(10):793-803.
- 256 3. Bova GS, Isaacs WB. Review of allelic loss and gain in prostate cancer. *World journal*
257 *of urology*. 1996; 14(5):338-46.
- 258 4. Dong JT, Suzuki H, Pin SS, Bova GS, Schalken JA, Isaacs WB, *et al*.
259 Down-regulation of the KAI1 metastasis suppressor gene during the progression of
260 human prostatic cancer infrequently involves gene mutation or allelic loss. *Cancer Res*.
261 1996; 56(19):4387-90.
- 262 5. Sugasawa K, Ng JM, Masutani C, Iwai S, van der Spek PJ, Eker AP, *et al*. Xeroderma
263 pigmentosum group C protein complex is the initiator of global genome nucleotide
264 excision repair. *Mol Cell*. 1998; 2(2):223-32.
- 265 6. Shen H, Sturgis EM, Khan SG, Qiao Y, Shahlavi T, Eicher SA, *et al*. An intronic poly
266 (AT) polymorphism of the DNA repair gene XPC and risk of squamous cell

- 267 carcinoma of the head and neck: a case-control study. *Cancer Res.* 2001;
268 61(8):3321-5.
- 269 7. Marin MS, Lopez-Cima MF, Garcia-Castro L, Pascual T, Marron MG, Tardon A. Poly
270 (AT) polymorphism in intron 11 of the XPC DNA repair gene enhances the risk of
271 lung cancer. *Cancer Epidemiol Biomarkers Prev.* 2004; 13(11 Pt 1):1788-93.
- 272 8. Sanyal S, De Verdier PJ, Steineck G, Larsson P, Onelov E, Hemminki K, *et al.*
273 Polymorphisms in XPD, XPC and the risk of death in patients with urinary bladder
274 neoplasms. *Acta oncologica (Stockholm, Sweden).* 2007; 46(1):31-41.
- 275 9. Vogel U, Overvad K, Wallin H, Tjonneland A, Nexø BA, Raaschou-Nielsen O.
276 Combinations of polymorphisms in XPD, XPC and XPA in relation to risk of lung
277 cancer. *Cancer Lett.* 2005; 222(1):67-74.
- 278 10. Sak SC, Barrett JH, Paul AB, Bishop DT, Kiltie AE. The polyAT, intronic IVS11-6
279 and Lys939Gln XPC polymorphisms are not associated with transitional cell
280 carcinoma of the bladder. *Br J Cancer.* 2005; 92(12):2262-5.
- 281 11. Gil J, Ramsey D, Stembalska A, Karpinski P, Pesz KA, Laczmanska I, *et al.* The C/A
282 polymorphism in intron 11 of the XPC gene plays a crucial role in the modulation of
283 an individual's susceptibility to sporadic colorectal cancer. *Mol Biol Rep.* 2011.
- 284 12. Kraemer KH, Lee MM, Andrews AD, Lambert WC. The role of sunlight and DNA
285 repair in melanoma and nonmelanoma skin cancer. The xeroderma pigmentosum
286 paradigm. *Arch Dermatol.* 1994; 130(8):1018-21.
- 287 13. [Polymorphism of DNA repair genes (XRCC1, XRCC3, XPC, XPD, XPA) in ethnic
288 groups from Republic of Bashkortostan]. *Genetika.* 2013; 49(8):1000-7.
- 289 14. Ahmad Aizat AA, Siti Nurfatimah MS, Aminudin MM, Ankathil R. XPC Lys939Gln
290 polymorphism, smoking and risk of sporadic colorectal cancer among Malaysians.
291 *World J Gastroenterol.* 2013; 19(23):3623-8.

- 292 15. Dong Z, Guo W, Zhou R, Wan L, Li Y, Wang N, *et al.* Polymorphisms of the DNA
293 repair gene XPA and XPC and its correlation with gastric cardiac adenocarcinoma in a
294 high incidence population in North China. *J Clin Gastroenterol.* 2008; 42(8):910-5.
- 295 16. Fontana L, Bosviel R, Delort L, Guy L, Chalabi N, Kwiatkowski F, *et al.* DNA repair
296 gene ERCC2, XPC, XRCC1, XRCC3 polymorphisms and associations with bladder
297 cancer risk in a French cohort. *Anticancer Res.* 2008; 28(3B):1853-6.
- 298 17. Jiang X, Zhou LT, Zhang SC, Chen K. XPC Polymorphism Increases Risk of
299 Digestive System Cancers: Current Evidence from A Meta-Analysis. *Chinese journal*
300 *of cancer research = Chung-kuo yen cheng yen chiu.* 2012; 24(3):181-9.
- 301 18. Jiao X, Ren J, Chen H, Ma J, Rao S, Huang K, *et al.* Ala499Val (C > T) and
302 Lys939Gln (A > C) polymorphisms of the XPC gene: their correlation with the risk of
303 primary gallbladder adenocarcinoma : a case-control study in China. *Carcinogenesis.*
304 2010.
- 305 19. Sanyal S, Festa F, Sakano S, Zhang Z, Steineck G, Norming U, *et al.* Polymorphisms
306 in DNA repair and metabolic genes in bladder cancer. *Carcinogenesis.* 2004;
307 25(5):729-34.
- 308 20. Sak SC, Barrett JH, Paul AB, Bishop DT, Kiltie AE. Comprehensive analysis of 22
309 XPC polymorphisms and bladder cancer risk. *Cancer Epidemiol Biomarkers Prev.*
310 2006; 15(12):2537-41.
- 311 21. Khan SG, Metter EJ, Tarone RE, Bohr VA, Grossman L, Hedayati M, *et al.* A new
312 xeroderma pigmentosum group C poly(AT) insertion/deletion polymorphism.
313 *Carcinogenesis.* 2000; 21(10):1821-5.
- 314 22. Khan SG, Muniz-Medina V, Shahlavi T, Baker CC, Inui H, Ueda T, *et al.* The human
315 XPC DNA repair gene: arrangement, splice site information content and influence of a

316 single nucleotide polymorphism in a splice acceptor site on alternative splicing and
317 function. *Nucleic Acids Res.* 2002; 30(16):3624-31.

318 23. Rondelli CM, El-Zein RA, Wickliffe JK, Etzel CJ, Abdel-Rahman SZ. A
319 comprehensive haplotype analysis of the XPC genomic sequence reveals a cluster of
320 genetic variants associated with sensitivity to tobacco-smoke mutagens. *Toxicol Sci.*
321 2010; 115(1):41-50.

322 24. Amos CI, Caporaso NE, Weston A. Host factors in lung cancer risk: a review of
323 interdisciplinary studies. *Cancer Epidemiol Biomarkers Prev.* 1992; 1(6):505-13.

324 25. Khoury MJ, Adams MJ, Jr., Flanders WD. An epidemiologic approach to ecogenetics.
325 *Am J Hum Genet.* 1988; 42(1):89-95.

326 26. Wang YC, Chen CY, Chen SK, Chang YY, Lin P. p53 codon 72 polymorphism in
327 Taiwanese lung cancer patients: association with lung cancer susceptibility and
328 prognosis. *Clin Cancer Res.* 1999; 5(1):129-34.

329 27. Jin X, Wu X, Roth JA, Amos CI, King TM, Branch C, *et al.* Higher lung cancer risk
330 for younger African-Americans with the Pro/Pro p53 genotype. *Carcinogenesis.* 1995;
331 16(9):2205-8.

332 28. Kuroda Y, Tsukino H, Nakao H, Imai H, Katoh T. p53 Codon 72 polymorphism and
333 urothelial cancer risk. *Cancer Lett.* 2003; 189(1):77-83.

334 29. Matullo G, Palli D, Peluso M, Guarrera S, Carturan S, Celentano E, *et al.* XRCC1,
335 XRCC3, XPD gene polymorphisms, smoking and (32)P-DNA adducts in a sample of
336 healthy subjects. *Carcinogenesis.* 2001; 22(9):1437-45.

337 30. Shen H, Spitz MR, Qiao Y, Guo Z, Wang LE, Bosken CH, *et al.* Smoking, DNA
338 repair capacity and risk of nonsmall cell lung cancer. *Int J Cancer.* 2003; 107(1):84-8.
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341 Tables

342

343 **Table1** Characteristics of prostate cancer cases and health controls.

344

	Cases	Controls
n	152	142
Age		
Mean±SD	71.68±8.97	70.19±10.86
Range	35-93	32-92
Smoking Status		
Non-Smoker	48	23
Smoker(light and heavy)	86	96*
Unknown	18	23

Two sided χ^2 test and Mann-Whitney where appropriate

*:P<0.01 Two-sided χ^2 test

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351 **Table 2** XPC Intron 11 C/A genotype frequency and distribution

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Genotype	Cases(n=152)	Controls(n=142)	Ajusted OR [†] (95% CI)	P [‡] value
C/C	39(25.7%)	57(40.1%)	1.00(Reference)	
C/A	81(53.3%)	62(43.7%)	1.91(1.13-3.24)	0.02
A/A	32(21.1%)	23(16.2%)	2.03(1.03-3.98)	0.04

†: Ajusted by age

‡: Two-sided χ^2 test

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367 **Table 3** XPC Intron 11 C/A genotype frequency and distribution divided into smoking status

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Smoking status	Cases(%)			Controls(%)			Ajusted OR [†] (95% CI)		P [‡] value		
	n	C/C	C/A	A/A	n	C/C	C/A	A/A			
Non-Smoker	48	11(22.9)	22(45.8)	15(31.3)	23	11(47.8)	10(43.5)	2(8.7)	1.00(reference)	7.70(1.38-42.88)	0.02
Light-Smoker(<35packs year)	40	14(35.0)	20(50.0)	6(15.0)	43	15(34.9)	21(48.8)	7(16.3)	1.00(reference)	0.86(0.24-3.31)	0.86
Heavy-Smoker(\geq 35packs year)	46	10(21.7)	29(63.0)	7(15.2)	53	21(39.6)	23(43.4)	9(17.0)	1.00(reference)	1.26(0.35-4.61)	0.44

[†]: Adjusted by age

[‡]: Two-sided χ^2 test

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